Proof-of-principle demonstration of a Mueller matrix decomposition method for polarized light tissue characterization in vivo

Michael F. G. Wood*
Nirmalya Ghosh*
Eduardo H. Moriyama
Brian C. Wilson
Ontario Cancer Institute
Division of Biophysics and Bioimaging
University Health Network
610 University Avenue
Toronto, Ontario M5G 2M9 Canada

I. Alex Vitkin†
Ontario Cancer Institute
Division of Biophysics and Bioimaging
University Health Network
610 University Avenue
Toronto, Ontario M5G 2M9 Canada

1 Introduction

Many tissue constituents, including collagen, muscle fibers, keratin, and glucose, interact with polarized light. Changes in tissue structure, in particular collagen content and organization, are associated with a number of pathologies. The ability to extract individual polarization properties from tissues has a number of biomedical applications, including measurements of tissue organization and anisotropy via birefringence, and the changes in these following various treatments, as well as determining concentrations of optical active molecules such as glucose via optical rotation. However, light scattering in the visible-near-infrared wavelength range complicates the extraction of important polarization information in tissue, due to the resulting depolarization and alteration of the incident polarized light. In addition, as several polarizing effects in tissue occur simultaneously, they must be individually decoupled and recovered from the overall measured polarization signals.

To perform quantitative polarimetry in tissue, a method of extracting individual polarization effects in the presence of scattering must be employed. We have previously reported a sensitive polarimetry platform that measures low polarization signals from scattering media, and employs a polar Mueller matrix decomposition method to decouple the combined effects from the experimentally obtained Mueller matrices. Presently, to demonstrate this approach in vivo, we report on in vivo measurements in a dorsal skin window chamber mouse model, using collagenase to induce changes in tissue structure and anisotropy (birefringence). The skin window chamber model is suitable for birefringence modulation via collagen alterations, as collagens are the main skin component, accounting for ~60% of the dry weight.

2 Materials and Methods

The polarization of light is well described through either Jones or Mueller matrix algebra. However, the latter allows for depolarization interactions and so is preferred for tissue polarimetry. In this approach, a four-element Stokes vector \( \mathbf{S} \) describes the intensity and polarization of the light beam. The transfer function of the Stokes vector for the light propagation through a medium that alters the polarization state is represented as a \( 4 \times 4 \) Mueller matrix \( \mathbf{M} \). The interaction with any medium is described by \( \mathbf{S}' = \mathbf{MS} \), where \( \mathbf{S} \) and \( \mathbf{S}' \) are the input and output Stokes vectors, respectively.

To measure the polarization effects in optically turbid samples, including tissues, we have developed a sensitive Stokes polarimeter system. Briefly, this employs polarization...
modulation using a photoelastic modulator (IS-90, Hinds Instruments Hillsboro, Oregon) and synchronous lock-in detection (SR830, Stanford Research Systems, Sunnyvale, California), allowing sensitive low-noise measurements of the Stokes vector of the light interacting with a turbid sample. By cycling the polarization of the incident beam (HeNe laser, 15 mW, λ=632.8 nm) using a linear polarizer and quarter-wave-plate combination, and measuring the output Stokes vectors, the Mueller matrix of the sample can be constructed. A schematic of the experimental system is shown in Fig. 1.

However, in a complex medium such as biological tissues, several polarization-altering effects occur simultaneously, and the resultant measured Mueller matrix reflects these in a coupled complicated manner. Thus, a method to extract the individual effects is required to quantify contributions of individual polarization effects, such as birefringence, optical activity, and depolarization. To accomplish this, we have developed a polar decomposition method suitable for use in highly scattering samples, where the measured matrix \( \mathbf{M} \) is decomposed into “basis” matrices representing the individual effects. In this approach, a depolarization matrix \( \mathbf{M}_\Delta \), a retardance matrix \( \mathbf{M}_R \), and a diattenuation matrix \( \mathbf{M}_D \) are employed, with \( \mathbf{M} = \mathbf{M}_\Delta \mathbf{M}_R \mathbf{M}_D \). From these decomposed matrices, useful polarization properties of interest can be decoupled and calculated. Specifically, the elements of \( \mathbf{M}_\Delta \) can be used to calculate the net depolarization coefficient (\( \Delta \)), the elements of \( \mathbf{M}_R \) can be used to calculate the linear retardance (birefringence \( \delta \)) and optical activity (\( \phi \)), and the elements of \( \mathbf{M}_D \) can be used to calculate the diattenuation (\( d \)). This decomposition process is summarized in Fig. 1.

We have previously validated this method theoretically with a polarization-sensitive Monte Carlo model and experimentally using tissue-simulating phantoms, and performed measurements on ex vivo tissue samples. To demonstrate its first use in vivo, we presently report measured changes in tissue structure using a dorsal skinfold window chamber mouse model. In this model, the inner skin layer of an athy-
mic nude mouse (NCRNU-M, Taconic, Hudson, New York) was removed from a 10-mm-diam region on the dorsal surface and a titanium saddle was sutured in place to hold the skin flap vertically, as shown in Fig. 1. A protective glass coverslip (145 ± 15 µm thick) was placed over the exposed tissue plane. Thus, a thin layer of tissue (∼500 µm) extends away from the animal, allowing for direct optical transmission measurements of polarized light through this layer, as shown in Fig. 1. This model enables accurate measurements in an in vivo setting, free of many of the challenges inherent in using fully 3-D tissue structures.

To induce changes in this skin preparation, 5 µl of collagenase (gelatinase) solution (Sigma-Aldrich, collagenase from Clostridium histolyticum) at a concentration of 1 mg ml⁻¹ was microinjected into one region of the tissue. Collagenase cleaves collagen fibers by breaking the peptide bonds connecting the monomer peptide units. Since collagen fibers are one of the primary sources of tissue birefringence and scattering, their destruction should change the polarization of the light propagating through this treated tissue region. Measurements of M were performed both in the region of collagenase injection and a distant control region, as shown in Fig. 1. Measurements were made with a 1-mm-diam laser beam before collagenase treatment and for 5 h postinjection at 30-min intervals, with an additional measurement at 24 h. An ink circle with a diameter of approximately 2 mm was marked around the control and treated regions so that the measurements could be repeated and correlated with subsequent histology. The treatment and measurements were carried out under general anesthesia induced by intraperitoneal injection of ketamine/xylazine (80- and 5-mg Kg⁻¹, respectively). Before each measurement, the animal was reanesthetized. Values for birefringence, depolarization, optical activity, and diattenuation were extracted from the experimentally derived Mueller matrices at each time point, through the prior polar decomposition approach. On completion of the measurements (24-h postcollagenase injection), the animal was sacrificed and the control and treated tissues were removed, sectioned laterally, and stained with hematoxylin and eosin (HE) for histological examination.

3 Results and Discussion

The derived birefringence and net depolarization values in both collagenase-treated and control regions are shown in Fig. 2. Values for optical activity and diattenuation were also extracted; however, these did not change appreciably with treatment and so the data are not shown. Histology (HE stain) of treated and control regions are shown in Fig. 3. As expected, injection of collagenase induced structural changes in the skin, seen as a loss of collagen fibers and the presence of inflammation and vasodilatation, which resulted in a more shortened path length than the overall path length. Using this relationship, the birefringence values in the treatment region prior to treatment and in the control region were calculated as \( B_n = 2.4 \times 10^{-4}\) and \( B_n = 2 \times 10^{-4}\), respectively. After treatment, the value decreased to \( B_n = 0.6 \times 10^{-4}\). These birefringence levels compare reasonably well with those found in the literature for tissue birefringence; typical values are in the range of \( 1 \times 10^{-5}\).

Fig. 2 (a) Birefringence and (b) depolarization before and as a function of time after collagenase injection in the treated and control regions. Symbols are experimentally derived values.
A decrease in the net depolarization coefficient $\Delta$ after treatment, from $-0.63$ to $-0.45$, was measured in the treated region and is plotted together with the values in the control region in Fig. 2(b), the latter again being essentially constant. This reduction in depolarization is also due to the destruction of the collagen fibers, since these represent one of the primary scattering structures in tissue. As with changes in birefringence, this is also confirmed by histology: see Fig. 3(b).

Using these changes in the depolarizing properties of the tissue, we were able to approximate the scattering coefficient of the tissue using our polarization-sensitive Monte Carlo simulation platform. Simulations were run with a 500-\(\mu\)m-thick slab geometry, where the absorption coefficient ($\mu_a$), scattering anisotropy ($g$), and birefringence were held constant at $\mu_a=2.2$ cm$^{-1}$, $g=0.9$, and $\Delta n=4 \times 10^{-4}$ for control/pre-treatment, while $\Delta n$ was set to $1 \times 10^{-4}$ for post-treatment (optical properties estimated for skin). The scattering coefficient was varied from $\mu_s=100$ to 250 cm$^{-1}$ in the simulations. Monte-Carlo-generated net depolarization values for those photons exiting the sample in the forward direction were calculated, from which the scattering coefficients corresponding to $\Delta=0.63$ and $\Delta=0.45$ were selected. Using this approach, $\mu_s$ was estimated as 182 cm$^{-1}$ for the pretreatment tissue and as 134 cm$^{-1}$ for the post-treatment tissue.

These results show the ability of the method to quantify changes in tissue structure using polarized light in vivo. The destruction of the collagen fibers in the extracellular matrix results in reduced tissue anisotropy and a reduction in scattering effects. These changes produce reduced tissue birefringence and depolarization, respectively, as measured via the experimental Mueller matrix, individually extracted through the polar decomposition method, and further quantified via polarization-sensitive Monte Carlo simulations. An important point to note is that, as seen in the upper-right photograph in Fig. 1, the treated region has markedly increased blood content compared to the control tissue, which would result in marked increase in the tissue absorption. Despite this, the Muller matrix decomposition technique is able to yield the underlying polarization parameters.

4 Conclusions

These results present the first in vivo demonstration of Mueller matrix decomposition-based polarimetric characterization of tissues. The interpretation of measured changes in values for birefringence and depolarization is consistent with histology results, and shows promise for the method’s ability to accurately quantify biologically relevant tissue parameters such as scattering and birefringence. This initial proof-of-principle study is being followed up by more extensive investigations of the use of this method in vivo, both for noninvasive measurements of glucose and for monitoring the response of infarcted myocardial tissues to stem-cell therapies. These studies will investigate whether similar measurements can be made in reflection geometries on bulk tissues, considering the additional complicating factors present with such geometries (such as sampling volume calculations and the effect of additional tissue layers). Clearly, there are many other potential applications, both in tissue diagnostics and in treatment response monitoring.

References