Specific immobilization of influenza A virus on GaAs (001) surface

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1 Introduction

The need to identify rapidly and with high sensitivity different viral/bacterial pathogens, fungi, and toxins is one of the major forces driving extensive research addressing the development of biosensors. The conventional detection of viruses has been carried out with cell culture,1 immunological methods,2–4 and molecular methods including the polymerase chain reaction.5 These techniques, however, require much time and expertise.6,7 The development of detection methods that are specific for targeted biomolecules and are easy to carry out is necessary for medical diagnostics, clinical analysis, or even field tests.8–10 Biosensors containing organic molecules offer promising solutions due to their speed, simplicity, and continuous monitoring capability.11 Such sensors could comprise antibodies immobilized via organic molecules on glass surface, e.g., of a microstructured fiber,12 the Au surface of a semiconductor surface plasmon resonance device,13 or a Si surface.14 Electrical properties of bacteria have also been taken advantage of in constructing an optoelectrical biosensor.15 Optical and electronic properties of III-V and II-VI semiconductor quantum well (QW) and quantum dot (QD) microstructures are also attractive for building biosensing devices because they can be used to detect miniscule perturbations of the semiconductor surface induced by selectively trapped biomolecules. For instance, the bright photoluminescence (PL) of a colloidal CdSe QD was investigated to develop fluorescent probes in sensing, imaging, immunoassay, and some other diagnostics applications.16–22 Recently, we proposed that templates of epitaxial QDs, such as InAs QDs in a GaAs matrix, could be used to construct a family of innovative biosensors with a significant potential to address the rapid detection of numerous pathogens in parallel.23 Our interest in GaAs is driven both by its strong PL

Abstract. In the quest for the development of an all-optical biosensor for rapid detection and typing of viral pathogens, we investigate biosensing architectures that take advantage of strong photoluminescence emission from III-V quantum semiconductors (QS). One of the key elements in the development of such a biosensor is the ability to attach various analytes to GaAs—a material of choice for capping III-V QS of our interest. We report on the study of biofunctionalization of GaAs (001) with polyethylene-glycol (PEG) thiols and the successful immobilization of influenza A virus. A diluted solution of biotinylated PEG thiols in OH-terminated PEG thiols is used to form a network of sites for the attachment of neutravidin. Biotinylated polyclonal influenza A antibodies are applied to investigate the process of the immobilization of inactivated influenza A virus. The successful immobilization is demonstrated using atomic force microscopy and fluorescence microscopy measurements. © 2009 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3251057]

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and its applications as a capping material for IR-emitting InAs QDs and some other quantum semiconductor microstructures. Consequently, the ability to immobilize specifically targeted biomolecules on the surface of GaAs is of high interest for the development of a proposed biosensor. We recently reported on the successful immobilization of avidin on the surface of GaAs.

In this paper, we report on the specific immobilization of inactivated influenza A virus using an architecture based on antibody and a self-assembled monolayer (SAM) of polyethylene-glycol (PEG) thiol deposited on the GaAs (001) surface.

2 Experimental Methods

2.1 Materials

Wafers of GaAs (001), series VW 10680-53 and 54, were bought from Wafer Technology Ltd. (Milton Keynes, United Kingdom). Polyclonal antibodies against the virus of influenza (H3N2) coupled with biotin or coupled with fluorescein isothiocyanate (FITC) were obtained from ViroStat, Inc. (Portland, Maine). Polyclonal antibodies against the hepatitis B and herpes simplex virus type 1 (HSV1) FITC conjugated were also obtained from ViroStat. PBS 10× (phosphate buffered saline, pH 7.4) was bought from Sigma (Oakville, Canada). Neutravidin was obtained from Molecular Probes (Invitrogen, Burlington, Canada). Biotinylated PEG (polyethylene glycol) thiol and OH-terminated PEG thiol were obtained from Prochimia Surfaces (Gdansk, Poland). The molecular structure of these thiols is shown in Fig. 1. Gamma ray inactivated viral particles of influenza A (rank 2, stock Texas 1/77) were bought from Microbix Bio-Systems (Toronto, Canada). The inactivation of the virus is achieved due to the damage of its RNA genome. In contrast to conventional chemical methods of inactivation that use formaldehyde, this method leaves the virus outer shell intact. Nominally anhydrous ethanol (98% v/v) was bought from Commercial Alcohols, Inc. (Brampton, Canada). To remove residual oxygen, a 250-mL flask filled with ethanol was purged for 4 h with a 3 SCFH high-purity nitrogen stream (<99%, Praxair Canada Inc.). OptiClear, a solvent designed to remove impurities present at the surface of optical or electric compounds, was obtained from National Diagnostics (Mississauga, Canada). Acetone was bought at ACP (Montréal, Canada); isopropanol (2-propanol) was obtained from Fisher Scientific (Ottawa, Canada), acetic acid (CH₃COOH) was obtained from Fisher Scientific, and ammonium hydroxide 28% (NH₄OH) was bought at Anachemia (Richmond, Canada). All the solvents were lab grade and all the products were used without additional purification.

2.2 Preparation of the Samples

A substrate of crystalline GaAs (001) was used to carry out the procedure of biofunctionalization. Samples of 4×4 mm, obtained by cleaving GaAs (001) wafers, were cleaned sequentially during 5 min in an ultrasonic bath of undiluted solutions of OptiClear, acetone, and isopropanol, similar to a previously described procedure. The samples were, thereaf-

After thiolation, the samples were rinsed with isopropanol to get rid of superfluous thiol molecules physically adsorbed to the substrate. Thereafter, they were immersed for 2 h in a PBS buffer containing a concentration of 200 μg/ml of neutravidin. This step was followed by rinsing of the samples with the PBS buffer and then with deionized water. For the immobilization of polyclonal antibodies against the influenza A virus, samples having been exposed to neutravidin were immersed for 2 h in a solution of biotinylated antibodies diluted (1:25) to a final concentration of 160 μg/ml in the PBS buffer. Once the incubation was completed, the samples were placed for 2 h in a solution containing 300 ng/ml (10 HA units/ml) of inactivated viral particles. This was followed by rinsing the samples with the PBS solution and storing them in PBS for characterization or future processing.

The procedure of biofunctionalization and the exposure of samples to the viral particles are schematically illustrated in Fig. 2. A series of samples biofunctionalized with antibodies against influenza A were prepared by exposing them to inactivated influenza A viruses. Additional samples were prepared by immersing the influenza A virus exposed samples for 2 h in FITC-conjugated antibodies against influenza A, hepatitis B, or herpes simplex virus type 1 (HSV1) diluted in PBS.
buffer. These samples, after rinsing with PBS and deionized water to remove residual salts from the surface, were used for fluorescence microscopy experiments.

2.3 Interface and Surface Characterization

2.3.1 Fourier transform infrared (FTIR) spectroscopy

To verify the process of PEG-thiol SAMs formation on the GaAs (001) surface, FTIR absorption was investigated in the region characteristic of the CH2 stretching vibrations (2800 to 3000 cm\(^{-1}\)). The spectra were collected using an FTIR Bruker Optics Hyperion 2000 microscope coupled with a spectrometer (Bruker Optics Tensor 27). The analyzed area was approximately 2 mm in diameter and the spectral resolution was 4 cm\(^{-1}\). A sample that was etched and incubated in ethanol was used to determine a baseline for the spectroscopy measurements.

2.3.2 Atomic force microscopy (AFM)

Surface morphology of processed samples was investigated using a Bioscope AFM (Veeco Metrology, Inc., California) operating in a contact mode. The biofunctionalized samples were never exposed to the atmosphere and their characterization was carried out in a PBS buffer solution. A MLCT-AUHW type tip (Veeco Metrology, Inc.) was used with a cantilever spring constant of 0.03 N/m. The AFM measurements of a reference GaAs sample (etched only) were carried out in an air ambient with contact mode using a Nanoscope E AFM (Veeco Metrology, Inc.).

2.3.3 Fluorescence microscopy

The fluorescence emitted by FITC fluorochrome (518 to 523 nm) attached to antibodies against influenza A was observed using fluorescence microscopy (Olympus IX71 inverted microscope with a DP71 digital camera). The excitation of the FITC fluorochrome was carried out with a blue light source emitting between 450 and 490 nm.

3 Results and Discussion

3.1 PEG Thiol SAM Formation on GaAs (001)

The formation of PEG thiol SAMs on the surface of GaAs (001) was investigated by examining the spectral location and intensity of the FTIR peaks corresponding to stretching vibrations of CH2 molecules. Figure 3 shows the FTIR spectra obtained for three different samples fabricated under nominally the same conditions. For each sample investigated, two peaks are observed near 2924 and 2853 cm\(^{-1}\), which corre-
respond respectively to the asymmetric and symmetric stretching mode vibrations of CH2. The lack of spectral definition makes the peak locations difficult to specify precisely, and moreover, the absorption intensity is more than 2 times weaker than that observed in dodecanethiol SAMs, which have the same \(n=11\) alkane number but that do form well-ordered SAMs.32,33 A recent study has shown that an additional IR enhancement factor of up to 6 times applies for the case of highly ordered \(n\)-alkane SAMs in transmission measurements.34 This carries the implication that SAMs having an increased degree of conformational disorder may belie the extent of the actual surface coverage by virtue of lower IR intensities observed, since the coverage and intensity will vary in a nonproportional manner. It is also expected that the PEG group will hinder ordering and surface coverage to some degree, but to just what extent is difficult to derive from our results for the reasons stated. More effort will result in a better characterization of the surface, but ultimately, the value of this interface will be determined by its functionality.

### 3.2 Surface Morphology and Biofunctionalization

The surface morphology of an etched GaAs (001) sample is shown by an AFM micrograph in Fig. 4(a). The root-mean-square roughness amplitude of the investigated 3.25-\(\times\)3.25-\(\mu\)m area is 0.3 nm. This value is expected for the high-quality GaAs (001) surface, and it is comparable to the results reported in literature.35 Figure 4(b) is an AFM micrograph of the GaAs (001) surface biofunctionalized with the influenza A antibody [as illustrated by the step represented in Fig. 2(d)]. The micrograph shows a granular microstructure composed of nanoparticles, each between 5 and 15 nm in diameter. These could represent neuravidin, the largest size biomolecule present in the studied sample. However, we have observed that samples exposed exclusively to the PBS buffer solution exhibited qualitatively a similar surface morphology to that represented in Fig. 4(b). This suggests that some PBS nanoparticles precipitate on the surface of samples studied by liquid-phase AFM, making it difficult to distinguish nanoparticles of different origin. An AFM micrograph of a fully biofunctionalized GaAs (001) sample that was exposed to the influenza A virus is shown in Fig. 4(c). A significantly different surface morphology in this case is characterized by the presence of large nanoparticles, typically 120 to 250 nm wide at the base and 50 to 100 nm tall. It is known that the influenza virus is a quasispherical object, approximately 100 to 120 nm in diameter.36,37 Therefore, it seems reasonable to assume that the large nanoparticles in Fig. 4(c) represent either individual or clustered influenza virus particles. The tip convolution effect makes the viral particles detected appear larger than 120 nm and a higher magnification experiment would have to be carried out to view details of the viral particle.38 The slightly reduced height of the viral particles observed in this experiment could be explained by the possible flattening occurring due to the pressure exercised by the AFM cantilever tip. Using AFM images, we have estimated that the average density of Influenza A particles covering the biofunctionalized GaAs (001) surface is approximately at 45 per 100 \(\mu\)m².

Fig. 4 AFM images and cross-sectional scans of (a) a reference GaAs (001) sample, (b) the GaAs (001) sample biofunctionalized with PEG thiols and neuravidin, and (c) the sample that following the functionalization with the influenza A antibody was exposed to a solution of inactivated influenza A virus.
3.3 Fluorescence Tests on Virus Exposed Samples

An example of the fluorescence microscopy image from the influenza A virus and to the FITC-conjugated antibodies against influenza A virus. Following this step, the samples were exposed to a 300 ng/ml solution comprising γ radiation-inactivated influenza A particles. With the AFM measurements, we identified the presence of viral nanoparticles on such samples. The surface density of immobilized particles was estimated as 45 per 100 μm². Using fluorescence microscopy, we were able to verify the nature of these nanoparticles. Indeed, bright fluorescence images were observed following the exposure of samples with immobilized influenza A virus to FITC-conjugated influenza A antibodies. Based on the fluorescence microscopy measurements, the density of influenza A particles specifically immobilized on the biofunctionalized GaAs (001) surface was estimated as at least 25 per 100 μm². To the best of our knowledge, these results are the first evidence of controlled immobilization of viral particles on GaAs. We consider this achievement to be an important step toward demonstration of a III-V semiconductor-based photonic biosensor.

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